

REVIEW

Open Access

# Proteostasis in pediatric pulmonary pathology

Silke Meiners\* and Korbinian Ballweg

## Abstract

Protein homeostasis describes the tight supervision of protein synthesis, correct protein maturation and folding, as well as the timely disposal of unwanted and damaged proteins by the ubiquitin-proteasome pathway or the lysosome-autophagy route. The cellular processes involved in preservation of protein homeostasis are collectively called proteostasis. Dysregulation of proteostasis is an emerging common pathomechanism for chronic lung diseases in the adult and aged patient. There is also rising evidence that impairment of protein homeostasis contributes to early sporadic disease onset in pediatric lung diseases beyond the well-known hereditary proteostasis disorders such as cystic fibrosis and alpha-1 antitrypsin (AAT) deficiency. Identifying the pathways that contribute to impaired proteostasis will provide new avenues for therapeutic interference with the pathogenesis of chronic lung diseases in the young and adult. Here, we introduce the concept of proteostasis and summarize available evidence on dysregulation of proteostasis pathways in pediatric and adult chronic lung diseases.

**Keywords:** Proteostasis; Conformational disorder; Autophagy; Proteasome; ER stress

## Introduction

Maintenance of a functional proteome and protein interaction network is essential for the survival of cells and organisms [1]. The cellular processes involved in preservation of this protein homeostasis are collectively called proteostasis. It covers all steps in the life of a protein, i.e., protein synthesis, correct protein maturation and folding, as well as the timely disposal of unwanted and damaged proteins by the ubiquitin-proteasome pathway or the lysosome-autophagy route (Figure 1). Dysfunctional proteostasis is emerging as a key and common pathomechanism for chronic lung diseases [2]. The range of lung disorders in which aberrant protein homeostasis has been implicated covers both pediatric and adult lung diseases that occur due to genetic, environmental, or idiopathic causes. The list includes chronic obstructive pulmonary disease (COPD) due to cigarette smoking or alpha-1 antitrypsin (AAT) deficiency, cystic fibrosis, idiopathic pulmonary fibrosis (IPF), pulmonary arterial hypertension (PAH), and some allergic airways diseases. Identifying the pathways of impaired proteostasis is central to interfere with the

pathogenesis of chronic lung diseases in the young and adult [3].

## Review

### Protein synthesis, folding, and function

Proteins are synthesized at the ribosome as a linear sequence of amino acids and fold into a complex three-dimensional structure that enables protein function [4]. Protein folding is usually coupled directly to protein translation but also takes place in subcellular compartments, such as the endoplasmic reticulum (ER) or mitochondria, and involves support by molecular chaperones, i.e., heat shock proteins (HSP) [5]. Molecular chaperones also regulate protein localization and protein-protein interactions, thereby contributing to functional protein networks [5]. Stress-induced protein modifications, lack of oligomeric assembly partners, or denaturation results in protein misfolding and challenges protein folding energetics and the cellular chaperone network [6]. Misfolded proteins expose hydrophobic surfaces that may trap other proteins to form protein aggregates, a process called proteotoxicity [6]. Molecular chaperones prevent protein aggregation by binding to these hydrophobic residues, thereby facilitating correct folding in an ATP-dependent manner [5]. Ensuring proper protein folding by molecular chaperone networks thus represents the first level of maintaining a functional proteostasis network. In case

\* Correspondence: silke.meiners@helmholtz-muenchen.de  
Comprehensive Pneumology Center (CPC), Member of the German Center for Lung Research (DZL), University Hospital, Ludwig-Maximilians-Universität, Asklepios Klinik Gauting und Helmholtz Zentrum München, Max-Lebsche-Platz 31, 81377 Munich, Germany

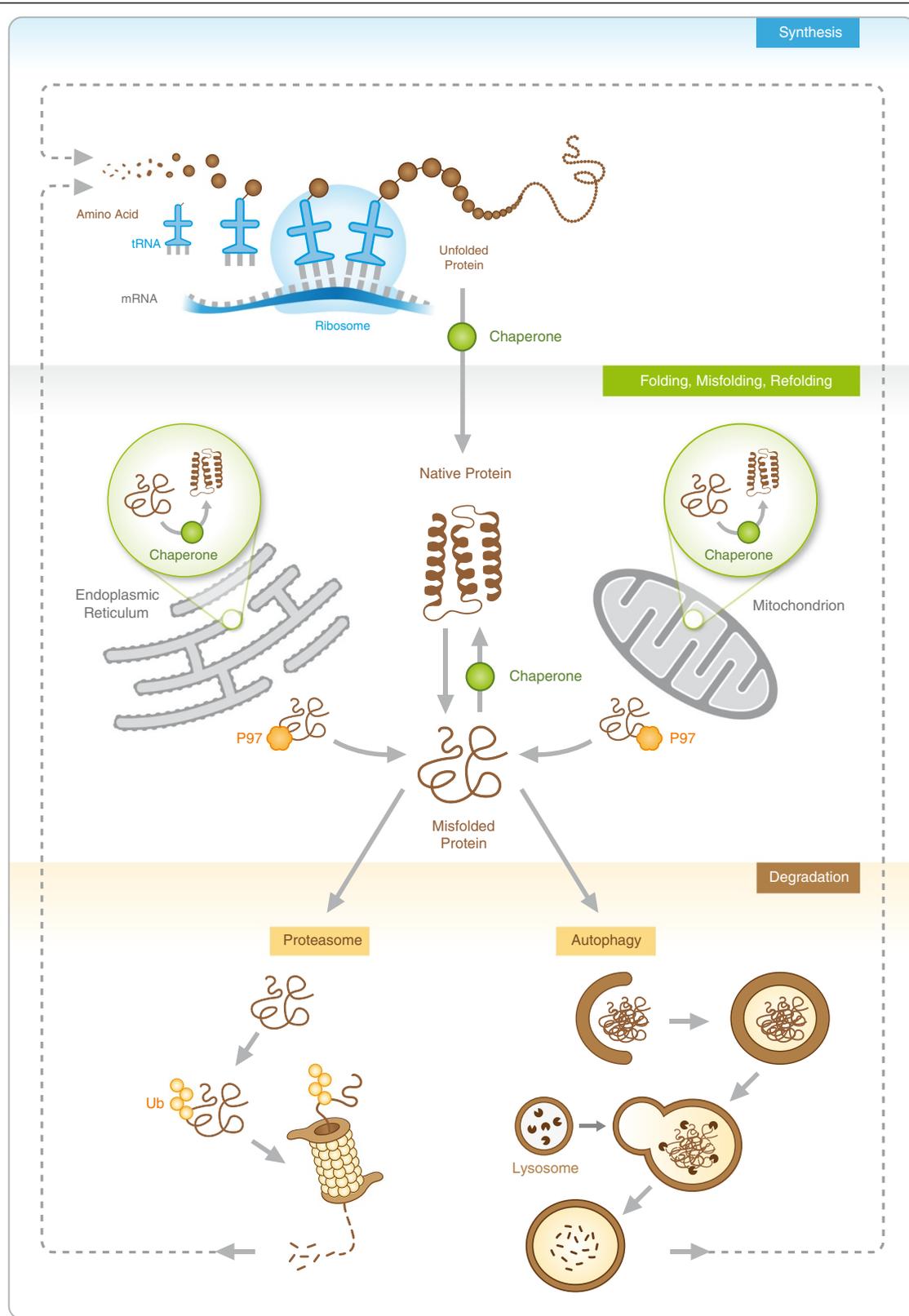


Figure 1 (See legend on next page.)

(See figure on previous page.)

**Figure 1 Life and death of proteins.** Proteins are synthesized as a linear amino acid chain at the ribosomes and fold into their native structure with the assistance of chaperones. Misfolded proteins in the cytosol or in cellular compartments, such as ER and mitochondria, are recognized by chaperones which support their refolding into the native structure. If misfolded proteins cannot be properly refolded, they are targeted for degradation mainly by the ubiquitin-proteasome system. For proteasomal degradation, proteins are tagged with a polyubiquitin chain and subsequently degraded by the proteasome into small peptides. Misfolded proteins of the ER or at the mitochondria are retro-translocated into the cytosol and transported to the proteasome for degradation with the help of VCP/p97. Aggregated proteins are engulfed by the autophagosome and degraded after fusion of lysosomes with the autophagosome. Degradation products of the proteasome and the autophagy pathway are mainly recycled as amino acids for protein synthesis.

chaperones are unable to successfully restore three-dimensional protein structure, misfolded proteins are targeted for proteasomal degradation with the help of specific protein quality ubiquitin ligases [7]. Insoluble protein aggregates are disposed by autophagy (Figure 1).

#### Protein degradation by the ubiquitin-proteasome or autophagy pathways

The ubiquitin-proteasome and the lysosomal autophagy pathways are the two main protein disposal systems in the cell. They perform not only housekeeping tasks of normal protein turnover but also protein quality control functions that ensure rapid disposal of misfolded and aggregated proteins.

The autophagy pathway comprises sequestration of cytoplasmic cargo in autophagic vesicles and subsequent degradation in the lysosome. Three different forms of autophagy can be distinguished: chaperone-mediated autophagy in which specific proteins are identified by chaperones and delivered directly to the lysosome, microautophagy which describes internalization of cytoplasmic contents by the lysosome, and macroautophagy for unspecific vesicle-mediated degradation via the lysosome [8]. Macroautophagy, often referred to simply as autophagy, allows disposal not only of bulky protein aggregates but also of dysfunctional organelles or parts thereof. For autophagy, the cargo is sequestered inside a double-membrane vesicle forming an autophagosome which then fuses with lysosomal vesicles to form the autolysosomes where the captured material is degraded by acid hydrolases [8]. Degradation products such as free amino and fatty acids are recycled, thereby constituting the prosurvival function of autophagy, e.g., in response to starvation [8]. On the molecular level, more than 30 autophagy-related genes (ATG) execute all autophagy steps [8]. Impaired nutrient supply as well as multiple forms of cellular stress induce autophagy by regulating expression and activation of ATG [9]. Moreover, autophagy also contributes to innate and adaptive immune surveillance and regulation of host/pathogen responses [10].

Degradation of specific cellular proteins is mainly taken over by the ubiquitin-proteasome system. Specificity of protein degradation is achieved by tagging of

degradation-prone proteins with polyubiquitin chains. Polyubiquitination can occur at different lysine residues of the ubiquitin molecules. Differentially linked ubiquitin chains have distinct cellular effects: Lysine (K)48-linked polyubiquitin chains, for example, target proteins for proteasomal degradation while K63 linkages are mainly associated with sorting of proteins for lysosomal degradation pathways [11]. Ubiquitination proceeds along an enzymatic cascade, in which ubiquitin is first activated by the ubiquitin-activating enzyme E1, conjugated by an E2 ubiquitin-conjugating enzyme, and finally covalently linked to the substrate by a specific E3 ubiquitin ligase [11,12]. Binding of further ubiquitin molecules to the previous one enables formation of a polyubiquitin chain which mediates degradation while modification of proteins with only one ubiquitin molecule mediates signaling [11]. Specific regulation of substrate turnover thus takes place on the level of ubiquitin-conjugating E3 ligases or - as a newly emerging theme - on the level of specific deubiquitinating enzymes, so-called DUBs [13]. The 26S proteasome is the proteolytic unit of the ubiquitin-proteasome system and specifically degrades polyubiquitinated proteins. It is composed of one barrel-shaped 20S catalytic core and one or two cap-like 19S regulatory complexes. While the 19S regulators facilitate binding and deubiquitination, as well as ATP-dependent unfolding of substrates, the 20S proteasome core particle hydrolyzes proteins with the help of three proteolytic active sites into small peptides. The three active sites of the proteasome have different cleavage specificities with  $\beta$ 1 preferably cleaving after acid residues,  $\beta$ 2 after basic residues, and  $\beta$ 5 after hydrophobic residues. The active sites are therefore also named caspase-like, trypsin-like, and chymotrypsin-like active sites, respectively [12]. In immune cells or upon stimulation with interferon- $\gamma$ , the three standard active subunits are exchanged by inducible subunits to form the immunoproteasome. Degradation products of the proteasome are not only used for recycling of amino acids but also exploited for major histocompatibility complex (MHC) class I antigen presentation to define the cell's self towards the immune system [14]. The controlled degradation of cellular proteins is essential for cellular growth, signaling, transcription, immune responses, and protein quality control and places

the ubiquitin-proteasome system at the heart of proteostasis [15,16].

### **Organelle-specific proteostasis**

Dysfunctional proteins arise in all cellular compartments such as the plasma membrane, cytosol, nucleus, ER, and mitochondria (Figure 1). Misfolded nuclear proteins are degraded by nuclear proteasomes or via nucleophagy [17]. In contrast, as the ER and mitochondria are not accessible for the proteasome, specialized pathways compensate for degradation of misfolded proteins in these compartments. With nearly one third of all proteins being secreted via the ER and requiring complex folding and glycosylation, this organelle is equipped with specialized ER-resident chaperones which recognize misfolded proteins and facilitate their ubiquitination by ER-associated ubiquitin ligases and subsequent transport into the cytosol for ubiquitin-mediated proteasomal degradation. This process is dependent on the AAA ATPase VCP/p97 and is termed ER-associated degradation pathway (ERAD) [18]. A similar pathway exists for mitochondrial proteins, i.e., mitochondria-associated degradation (MAD) which involves disposal of mitochondrial proteins - mainly from the outer membrane - via a VCP/p97 and proteasome-dependent pathway [19].

In addition, misfolded proteins in these compartments trigger a compartment-specific unfolded protein response (UPR) which aims to adjust the capacity of folding and disposal in the organelle [20,21]. Sequestering of chaperones by misfolded proteins in the ER is rapidly sensed by three sensor proteins: IRE1, PERK, and ATF6, and launches the three-armed adaptive stress response of the ER. While activation of PERK attenuates protein translation thereby decreasing protein reloading of the ER, IRE1 and ATF6 concertedly activate UPR-specific transcription to increase the ER folding capacity and function. When they fail, however, e.g., upon severe or chronic ER stress, apoptosis is induced [22]. Accumulation of misfolded proteins in the mitochondrial matrix impairs the import of the transcription factor ATF5-1 into mitochondria but triggers its translocation to the nucleus to induce expression of mitochondrial proteases and chaperones [21]. A distinct mitochondrial unfolded protein response exists also for the intermembrane space [23]. Organelle proteostasis also involves bulk degradation of whole organelles via the autophagy pathway such as nucleophagy or mitophagy or by autophagic disposal of parts of the dysfunctional ER [17,24].

### **Proteostasis in chronic lung diseases**

Dysfunctional proteostasis is a common pathomechanism for both, pediatric and adult, chronic lung diseases that either are caused by inherited protein dysfunction or involve sporadic impairment of protein homeostasis

[2]. In this section, we will summarize available knowledge on proteostasis in the young and immature as well as in the adult and aged lung.

### **Hereditary dysfunction of proteostasis**

In the lung, the two major inherited genetic disorders, cystic fibrosis and AAT deficiency, are conformational disorders that are caused by the hereditary expression of mutant alleles of the cystic fibrosis transmembrane conductance regulator (CFTR) and AAT, respectively [25]. In addition, mutations of the surfactant proteins (SP) C, B, and A have been identified in familial cases of IPF [3,26]. All of these mutations pose a serious challenge for the proteostasis network as they result not only in impaired function of the respective protein but also in ER stress, dysregulated proteasome, and altered autophagy function [25-27]. The deltaF508CFTR mutant - the most predominant CFTR mutation in cystic fibrosis patients - is characterized by a misfolded CFTR protein which is rapidly degraded by the proteasome and thus fails to translocate to the plasma membrane [28]. Consequently, modulating the proteostasis network to stabilize folding and trafficking of this CFTR mutant was suggested as a beneficial therapeutic approach in CF [29]. Loss of functional CFTR has further been associated with defective autophagy and accumulation of aggregated and polyubiquitinated proteins. Restoration of autophagy increased CFTR trafficking to the cell membrane [15,30]. There is also evidence for enhanced ER stress in CF patients as summarized elsewhere [25], and inactivation of the XBP-1 pathway decreased inflammatory cytokine production in a model of inflamed CF airway linking ER stress to inflammation [31]. Likewise, in AAT deficiency, misfolded AAT variants fail to exit the ER and are targeted for ERAD. Some AAT variants polymerize and are degraded via autophagy [25]. Thereby, misfolded AAT accumulates in hepatocytes and cannot be secreted in the bloodstream, which leads to unopposed protease activity in the lung [30]. Activation of autophagy prevents aggregate toxicity in hepatocytes and reversed liver pathology in a mouse model of AAT deficiency [30]. Misfolded AAT primarily leads to proteostasis imbalance and ER stress mainly in the liver while the lung phenotype manifests later in life due to unopposed protease activation [25]. However, as aggregates of mutant AAT were also detected, e.g., in alveolar macrophages, enhanced aggregate clearance by autophagy might also be beneficial in the lung [30]. Similarly, most of the known mutations of the SP-C and SP-A genes result in abnormal processing, folding, and accumulation of the mutant protein in the ER, possibly triggering ER stress in alveolar epithelial type II cells (AECII). Subsequent induction of apoptosis in the alveolar epithelium may then initiate fibrotic tissue remodeling and chronic inflammatory responses

[26,32]. In addition to ER stress, dysfunction of the proteasome and autophagy pathways has been shown for some of the SP-C mutants *in vitro* [33]. The rare hereditary disorder Hermansky-Pudlak syndrome is also characterized by disturbed proteostasis, namely impaired biogenesis of lysosome-related organelles, which contributes to continuous damage and apoptosis of AECII and early onset of pulmonary fibrosis [34].

#### **Sporadic dysfunction of proteostasis**

In addition to hereditary conformational disorders of the lung, proteostasis is challenged by exposure of the lung to noxious particles, chemicals, allergens, and pathogens which starts early in life and continues throughout life, thereby contributing to age-related chronic lung diseases [27,35].

#### **Impaired proteostasis in the young and immature lung**

Mechanical ventilation and oxygen supply are often used as a treatment in preterm-born infants. While life-saving, both, mechanical hyperinflation and hyperoxia, damage the immature lung, thus contributing to development of chronic lung diseases such as bronchopulmonary dysplasia (BPD) which persists during childhood and adulthood [36]. Among others, mechanical ventilation and hyperoxia induce formation of reactive oxygen species (ROS) which challenge the proteostasis network of the immature lung due to oxidative protein modifications and protein folding stress [37,38]. Thus, exposure of newborn mice to hyperoxia induced expression of ER stress markers including CHOP that may contribute to increased apoptosis [39]. Furthermore, hyperoxia altered ER morphology and eIF2 $\alpha$  phosphorylation but did not affect other UPR pathways in a newborn rat model [40]. Whether mechanical ventilation and hyperoxia also trigger autophagy or proteasome dysfunction in infants is largely unknown. In adult mice lungs, however, autophagy is increased upon mechanical ventilation and contributes to cellular damage [41]. Furthermore, hyperoxia induced autophagosome formation and expression of autophagic proteins in bronchial epithelial cells and in mice. Enhanced expression of the autophagic protein LC3B-II decreased apoptosis in hyperoxia-treated cells by regulating the extrinsic apoptosis pathway [42]. In hyperoxia-treated adult mice, elevated levels of PINK1 helped to maintain mitochondrial homeostasis by inducing autophagy of damaged mitochondria. In this study, higher activity of the lung proteasome was observed as well [43]. Hyperoxia also augmented ubiquitination and expression of the chaperone HSP70 in bronchial epithelial cells from adult patients treated with >95% oxygen [44]. In contrast, HSP27 expression was decreased in hyperoxia-treated newborn rats and in a human alveolar epithelial cell line, A549 cells, while overexpression of

HSP27 in A549 cells attenuated hyperoxia-induced cell death [45]. In summary, there is rising evidence for dysfunctional proteostasis in the ER and cytosol in response to mechanical ventilation and/or hyperoxia in the adult and also immature lung.

Asthma is the most common respiratory lung disease in children affecting more than 10% of children between 2 and 15 years of age [46]. Recent studies support the concept of asthma, although primarily an inflammatory disease, as a disease initiated by recurrent dysfunction of the airway epithelium which modulates inflammatory responses [47]. Accumulating evidence suggests a key role for oxidative events and protein damage in the pathogenesis of bronchial asthma [48,49]. Accordingly, ER stress markers were found to be increased in blood cells and bronchoalveolar lavage (BAL) from asthmatic patients and in the lung tissue of ovalbumin-challenged mice. Furthermore, administration of chemical chaperones reduced experimental asthma and inflammation in mice [50]. This effect was mainly mediated by attenuating ER stress-induced NF $\kappa$ B activation [50]. As NF $\kappa$ B activation can also be attenuated by inhibition of the proteasome, proteasome inhibitors have also been suggested as potential therapeutics in asthma. However, although proteasome inhibitors had some beneficial anti-inflammatory effects, they were unable to ameliorate chronic asthma in mice [16]. In addition, autophagy was shown to be augmented in asthmatic patients: Expression of Atg5 was increased in nasal mucosal cells of children with acute asthma and was associated with a single nucleotide polymorphism (SNP) in the Atg5 promoter [51]. A second SNP in the ATG5 gene was also associated with the enhanced formation of autophagosomes [52]. Thus, some initial evidence implicates an involvement of autophagy in asthma, but the molecular mechanisms remain to be elucidated.

Another aspect of how impaired proteostasis may affect development of asthma relates to the role of the proteasome in MHC class I antigen presentation: As the peptide products of proteasomal protein degradation may be used for MHC class I antigen presentation, proteasome function is an essential part of the host response towards intracellular infections by activating CD8 $^{+}$  T cell-mediated adaptive immunity against viral infections [14]. Only one study has addressed the putative involvement of the immunoproteasomes in asthma development using knockout mice for the immunoproteasome subunit LMP7. LMP7-deficient animals mounted a strong Th2 response during ovalbumin-induced but not house dust mite-induced acute asthma [53]. The role of immunoproteasome-dependent immune responses to virus infection in asthma has not been investigated so far. In general, the contribution of proteostasis systems to sporadic lung disease in infants and children is only

beginning to be unraveled. Recent evidence shows that protein homeostasis is dysregulated in the development of BPD and asthma. However, to what extent the different proteostasis pathways contribute to these pediatric diseases, how they interact or compensate for each other, and whether they represent a feasible therapeutic target await further investigation.

**Deficient proteostasis in the adult and aged lung** Impaired proteostasis has been identified as a key and common pathomechanism for sporadic chronic lung diseases as reviewed extensively elsewhere [2,15,16,30]. Thus, we will only briefly outline the basic findings and would like to refer the reader to the details as given in the above cited reviews.

For sporadic chronic lung diseases in the adult, cigarette smoke remains the main risk factor. As such, cigarette smoke exposure of the lung represents a paradigm for environmental insults that drive pathogenesis of chronic lung diseases. Among the more than 4,700 chemicals of cigarette smoke, reactive compounds oxidatively modify cellular proteins, thereby seriously challenging the proteostasis network of the cell. Several proteins such as secretory proteins, histone-modifying enzymes, signaling mediators, and transcriptional regulators have been shown to be oxidatively modified by cigarette smoke, adding to cellular dysfunction and pathogenesis of chronic lung diseases [54,55]. In agreement with the increased burden of misfolded proteins, e.g., upon cigarette smoke exposure, protein quality control pathways are mainly activated in adult chronic lung diseases. Induction of autophagy is a prominent feature in the lungs of patients with COPD as well as in mice chronically exposed to cigarette smoke [56]. Autophagy is also increased in PAH tissue, and mice lacking the autophagic protein LC3B showed exaggerated features of PAH in a hyperoxia model [30,57]. These findings together with the observation of fragmented mitochondrial morphology in pulmonary smooth muscle cells of PAH patients raised some speculation on a pathogenetic contribution of mitophagy in PAH [58]. However, activation of mitophagy in PAH could not be confirmed experimentally. Conflicting data also exist for the role of mitochondrial quality control in COPD. In IPF, autophagy pathways are not activated but rather impaired [59]. A recent study as well as unpublished data from our lab suggests that proteostasis in IPF might be rather regulated on the level of 26S proteasome activation [3,60]. For COPD, conflicting data exist on proteasome function in the lung: While Malhotra et al. observed a decline in proteasome expression and activity in the lungs of COPD patients which correlated with progressive disease severity [61], Baker et al. did not observe any decline in proteasome activity in end-stage COPD tissue [60]. Induction of ER stress in the lung has been observed in response to

cigarette smoke exposure as well as in pulmonary fibrosis [62]. In transgenic mouse models, induction of ER stress in the alveolar epithelium predisposes to enhanced lung fibrosis after treatment with bleomycin [26]. Furthermore, ER stress markers are observed in pulmonary hypertension, and treatment of rats with the modulator of the unfolded protein response salubrinal attenuated experimental PAH [63].

Protein quality control pathways are not only activated as an adaptive response to altered proteostasis, but they may also be direct targets of noxious environmental exposures as recently shown for the proteasome. Acute exposure to cigarette smoke directly inhibits the catalytic activity of purified proteasomes and proteasome activity of bronchial and alveolar epithelial cells as well as in mouse lungs [64,65]. Oxidative impairment of the ER-resident protein disulfide isomerase by cigarette smoke also expands protein misfolding in the ER compartment [54]. Such impairment of protein quality control further exacerbates the detrimental effects of noxious exposures of the lung and may amplify progression of chronic lung diseases. In line with this notion, proteostasis has been shown to decline with aging [66]. As a consequence of aging, misfolded proteins accumulate over time and proteostasis pathways become functionally impaired in response to persistent or repeated environmental exposures, thereby amplifying protein damage [6]. Age-related impairment of proteostasis has been observed on all levels including diminished chaperone availability, altered autophagy regulation, impaired proteasome function, and activation of unfolded protein responses in the ER and mitochondria [66-68]. Of note, only very few studies analyzed the impact of an aged proteostasis network on the development or progression of chronic respiratory diseases using animal models [69]. Further research will be required to delineate the contribution of single proteostasis pathways to distinct age-related chronic lung diseases.

## Conclusions

Dysregulation of proteostasis is an emerging common pathomechanism for chronic lung diseases in the adult and aged patient. There is also rising evidence that sporadic impairment of protein homeostasis contributes to early disease onset in pediatric lung disorders beyond the well-known hereditary proteostasis disorders such as cystic fibrosis and AAT deficiency. Maintaining protein homeostasis in the lung is particularly relevant in any condition that leads to sustained generation of ROS, e.g., mechanical ventilation, hyperoxia, and exposure to noxious particles and gases. In particular, compartment-specific dysregulation of proteostasis such as the unfolded protein response of the ER and mitochondria and altered proteasome function are emerging as novel aspects of dysregulated proteostasis in the diseased lung. The interplay of the

proteostasis network with other essential cellular functions such as cilia and vesicle transport is only beginning to be unraveled [3]. In particular, the close interaction of protein homeostasis with immune response pathways is a largely neglected aspect of how impaired proteostasis within a single cell outreaches its neighborhood: Proteasome function is essential for MHC class I-mediated adaptive immunity against viral and intracellular infections [14]. In addition, autophagy is central for innate immune surveillance of pathogens and also affects MHC class II presentation [70]. Moreover, proper function of the ER is required to ensure expression of immune relevant surface molecules and secretion of cytokines [71].

This adds a novel aspect to the concept of modulating proteostasis for disease intervention as proposed by Bouche-careilh and Balch [27] and may stimulate further research on dysregulated proteostasis in the young and adult lung.

#### Abbreviations

AAT: alpha-1 antitrypsin; AECL: alveolar epithelial type II cells; ATG: autophagy-related genes; BAL: bronchoalveolar lavage; BPD: bronchopulmonary dysplasia; CFTR: cystic fibrosis transmembrane conductance regulator; COPD: chronic obstructive pulmonary disease; ER: endoplasmic reticulum; ERAD: ER-associated degradation; IPF: idiopathic pulmonary fibrosis; MAD: mitochondrial-associated degradation; MHC: major histocompatibility complex; PAH: pulmonary arterial hypertension; ROS: reactive oxygen species; SP: surfactant protein; UPR: unfolded protein response.

#### Competing interests

The authors declare that they have no competing interests.

#### Authors' contributions

SM and KB wrote the review and edited the text. Both authors read and approved the final manuscript.

Received: 1 August 2014 Accepted: 19 November 2014

Published online: 29 December 2014

#### References

1. Powers ET, Balch WE (2013) Diversity in the origins of proteostasis networks – a driver for protein function in evolution. *Nat Rev Mol Cell Biol* 14:237–248, doi:10.1038/nrm3542
2. Balch WE, Sznajder JI, Budinger S, Finley D, Laposky AD, Cuervo AM, Benjamin IJ, Barreiro E, Morimoto RI, Postow L, Weissman AM, Gail D, Banks-Schlegel S, Croxton T, Gan W (2013) NHLBI Workshop: Malfolded protein structure and proteostasis in lung diseases. *Am J Respir Crit Care Med* 130913130718005. doi:10.1164/rccm.201306-1164WS
3. Meiners S, Green CM (2014) Protein quality control in lung disease: it's all about cloud networking. *Eur Respir J*. doi:10.1183/09031936.00105214
4. Kim YE, Hipp MS, Bracher A, Hayer-Hartl M, Hartl FU (2013) Molecular chaperone functions in protein folding and proteostasis. *Annu Rev Biochem* 82:323–355, doi:10.1146/annurev-biochem-060208-092442
5. Hartl FU, Bracher A, Hayer-Hartl M (2011) Molecular chaperones in protein folding and proteostasis. *Nature* 475:324–332, doi:10.1038/nature10317
6. Morimoto RI (2008) Proteotoxic stress and inducible chaperone networks in neurodegenerative disease and aging. *Genes Dev* 22:1427–1438, doi:10.1101/gad.1657108
7. Wickner S (1999) Posttranslational quality control: folding, refolding, and degrading proteins. *Science* 286:1888–1893, doi:10.1126/science.286.5446.1888
8. He C, Klionsky DJ (2009) Regulation mechanisms and signaling pathways of autophagy. *Annu Rev Genet* 43:67–93, doi:10.1146/annurev-genet-102808-114910
9. Kroemer G, Mariño G, Levine B (2010) Autophagy and the integrated stress response. *Mol Cell* 40:280–293, doi:10.1016/j.molcel.2010.09.023
10. Crotzer VL, Blum JS (2010) Autophagy and adaptive immunity: autophagy and immunity. *Immunology*. doi:10.1111/j.1365-2567.2010.03321.x
11. Komander D, Rape M (2012) The ubiquitin code. *Annu Rev Biochem* 81:203–229, doi:10.1146/annurev-biochem-060310-170328
12. Finley D (2009) Recognition and processing of ubiquitin-protein conjugates by the proteasome. *Annu Rev Biochem* 78:477–513, doi:10.1146/annurev-biochem.78.081507.101607
13. Bedford L, Lowe J, Dick LR, Mayer RJ, Brownell JE (2011) Ubiquitin-like protein conjugation and the ubiquitin-proteasome system as drug targets. *Nat Rev Drug Discov* 10:29–46, doi:10.1038/nrd3321
14. Kloetzel PM, Ossendorf F (2004) Proteasome and peptidase function in MHC-class-I-mediated antigen presentation. *Curr Opin Immunol* 16:76–81
15. Meiners S, Eickelberg O (2012) What shall we do with the damaged proteins in lung disease? Ask the proteasome! *Eur Respir J* 40:1260–1268, doi:10.1183/09031936.00208511
16. Meiners S, Keller IE, Semren N, Caniard A (2014) Regulation of the proteasome: evaluating the lung proteasome as a new therapeutic target. *Antioxid Redox Signal* 140314142218004. doi:10.1089/ars.2013.5798
17. Mijaljica D, Devenish RJ (2013) Nucleophagy at a glance. *J Cell Sci* 126:4325–4330, doi:10.1242/jcs.133090
18. Claessen JHL, Kundrat L, Ploegh HL (2012) Protein quality control in the ER: balancing the ubiquitin checkbook. *Trends Cell Biol* 22:22–32, doi:10.1016/j.tcb.2011.09.010
19. Karbowski M, Youle RJ (2011) Regulating mitochondrial outer membrane proteins by ubiquitination and proteasomal degradation. *Curr Opin Cell Biol* 23:476–482, doi:10.1016/j.jceb.2011.05.007
20. Ron D, Walter P (2007) Signal integration in the endoplasmic reticulum unfolded protein response. *Nat Rev Mol Cell Biol* 8:519–529, doi:10.1038/nrm2199
21. Pellegrino MW (2013) Nargund AM, Haynes CM Signaling the mitochondrial unfolded protein response. *Biochim Biophys Acta BBA - Mol Cell Res*. doi:10.1016/j.bbamcr.2012.02.019
22. Shore GC, Papa FR, Oakes SA (2011) Signaling cell death from the endoplasmic reticulum stress response. *Curr Opin Cell Biol* 23:143–149, doi:10.1016/j.jceb.2010.11.003
23. Papa L, Germain D (2011) Estrogen receptor mediates a distinct mitochondrial unfolded protein response. *J Cell Sci* 124:1396–1402, doi:10.1242/jcs.078220
24. Rambold AS, Lippincott-Schwartz J (2011) Mechanisms of mitochondria and autophagy crosstalk. *Cell Cycle Georget Tex* 10:4032–4038, doi:10.4161/cc.10.23.18384
25. McElvaney NG, Greene CM (2012) Mechanisms of protein misfolding in conformational lung diseases. *Curr Mol Med* 12:850–859
26. Tanjore H, Blackwell TS, Lawson WE (2012) Emerging evidence for endoplasmic reticulum stress in the pathogenesis of idiopathic pulmonary fibrosis. *AJP Lung Cell Mol Physiol* 302:L721–L729, doi:10.1152/ajplung.00410.2011
27. Bouche-careilh M, Balch WE (2011) Proteostasis: a new therapeutic paradigm for pulmonary disease. *Proc Am Thorac Soc* 8:189–195, doi:10.1513/pats.201008-055MS
28. Turnbull EL, Rosser MFN, Cyr DM (2007) The role of the UPS in cystic fibrosis. *BMC Biochem* 8(Suppl 1):S11, doi:10.1186/1471-2091-8-S1-S11
29. Hetz C, Chevet E, Harding HP (2013) Targeting the unfolded protein response in disease. *Nat Rev Drug Discov* 12:703–719, doi:10.1038/nrd3976
30. Mizumura K, Cloonan SM, Haspel JA, Choi AMK (2012) The emerging importance of autophagy in pulmonary diseases. *Chest J* 142:1289, doi:10.1378/chest.12-0809
31. Ribeiro CMP, Boucher RC (2010) Role of endoplasmic reticulum stress in cystic fibrosis-related airway inflammatory responses. *Proc Am Thorac Soc* 7:387–394, doi:10.1513/pats.201001-017AW
32. Lawson WE, Grant SW, Ambrosini V, Womble KE, Dawson EP, Lane KB, Markin C, Renzoni E, Lympany P, Thomas AQ, Roldan J, Scott TA, Blackwell TS, Phillips JA 3rd, Loyd JE, du Bois RM (2004) Genetic mutations in surfactant protein C are a rare cause of sporadic cases of IPF. *Thorax* 59:977–980, doi:10.1136/thx.2004.026336
33. Mulugeta S (2005) A surfactant protein C precursor protein BRICHOS domain mutation causes endoplasmic reticulum stress, proteasome dysfunction, and caspase 3 activation. *Am J Respir Cell Mol Biol* 32:521–530, doi:10.1165/rcmb.2005-0009OC
34. Mahavadi P, Guenther A, Gochoico BR (2012) Hermansky-Pudlak syndrome interstitial pneumonia: it's the epithelium, stupid! *Am J Respir Crit Care Med* 186:939–940, doi:10.1164/rccm.201210-1771ED

35. Selman M, Pardo A (2014) Revealing the pathogenic and aging-related mechanisms of the enigmatic idiopathic pulmonary fibrosis. an integral model. *Am J Respir Crit Care Med* 189:1161–1172, doi:10.1164/rccm.201312-2221PP
36. Hilgendorff A, Reiss I, Ehrhardt H, Eickelberg O, Alvira CM (2013) Chronic lung disease in the preterm infant: lessons learned from animal models. *Am J Respir Cell Mol Biol* 130911135746008, doi:10.1165/rcmb.2013-0014TR
37. Berkelhamer SK, Kim GA, Radder JE, Wedgwood S, Czech L, Steinhorn RH, Schumacker PT (2013) Developmental differences in hyperoxia-induced oxidative stress and cellular responses in the murine lung. *Free Radic Biol Med* 61:51–60, doi:10.1016/j.freeradbiomed.2013.03.003
38. Birukov KG (2009) Cyclic stretch, reactive oxygen species, and vascular remodeling. *Antioxid Redox Signal* 11:1651–1667, doi:10.1089/ars.2008.2390
39. Choo-Wing R, Syed MA, Harjith A, Bowen B, Pryhuber G, Janz C, Andersson S, Homer RJ, Bhandari V (2013) Hyperoxia and interferon- $\gamma$ -induced injury in developing lungs occur via cyclooxygenase-2 and the endoplasmic reticulum stress-dependent pathway. *Am J Respir Cell Mol Biol* 48:749–757, doi:10.1165/rcmb.2012-0381OC
40. Konsavage WM, Zhang L, Wu Y, Shenberger JS (2012) Hyperoxia-induced activation of the integrated stress response in the newborn rat lung. *AJP Lung Cell Mol Physiol* 302:L27–L35, doi:10.1152/ajplung.00174.2011
41. Lopez-Alonso I, Aguirre A, Gonzalez-Lopez A, Fernandez AF, Amado-Rodriguez L, Astudillo A, Batalla-Solis E, Albaiceta GM (2013) Impairment of autophagy decreases ventilator-induced lung injury by blockade of the NF- $\kappa$ B pathway. *AJP Lung Cell Mol Physiol* 304:L844–L852, doi:10.1152/ajplung.00422.2012
42. Tanaka A, Jin Y, Lee SJ, Zhang M, Kim HP, Stolz DB, Ryter SW, Choi AM (2012) Hyperoxia-induced LC3B interacts with the Fas apoptotic pathway in epithelial cell death. *Am J Respir Cell Mol Biol* 46:507–514, doi:10.1165/rcmb.2009-0415OC
43. Zhang Y, Sauler M, Shinn AS, Gong H, Haslip M, Shan P, Mannam P, Lee PJ (2014) Endothelial PINK1 mediates the protective effects of NLRP3 deficiency during lethal oxidant injury. *J Immunol* 192:5296–5304, doi:10.4049/jimmunol.1400653
44. Chambellan A, Cruickshank PJ, McKenzie P, Cannady SB, Szabo K, Comhair SA, Erzurum SC (2006) Gene expression profile of human airway epithelium induced by hyperoxia *in vivo*. *Am J Respir Cell Mol Biol* 35:424–435, doi:10.1165/rcmb.2005-0251OC
45. Shao L, Perez RE, Gerthoffer WT, Truong WE, Xu D (2009) Heat shock protein 27 protects lung epithelial cells from hyperoxia-induced apoptotic cell death. *Pediatr Res* 65:328–333, doi:10.1203/PDR.0b013e3181961a51
46. British Thoracic Society (2006) The burden of lung disease. British Thoracic Society, London UK
47. Holgate ST (2010) Has the time come to rethink the pathogenesis of asthma? *Curr Opin Allergy Clin Immunol* 10:48–53, doi:10.1097/ACI.0b013e3283347be5
48. Sugiura H, Ichinose M (2008) Oxidative and nitrative stress in bronchial asthma. *Antioxid Redox Signal* 10:785–798, doi:10.1089/ars.2007.1937
49. Zuo L, Ottenbaker NP, Rose BA, Salisbury KS (2013) Molecular mechanisms of reactive oxygen species-related pulmonary inflammation and asthma. *Mol Immunol* 56:57–63, doi:10.1016/j.molimm.2013.04.002
50. Kim SR, Kim DI, Kang MR, Lee KS, Park SY, Jeong JS, Lee YC (2013) Endoplasmic reticulum stress influences bronchial asthma pathogenesis by modulating nuclear factor  $\kappa$ B activation. *J Allergy Clin Immunol* 132:1397–1408.e11, doi:10.1016/j.jaci.2013.08.041
51. Martin LJ, Gupta J, Jyothula SS, Butsch Kovacic M, Biagini Myers JM, Patterson TL, Erickson MB, He H, Gibson AM, Baye TM, Amirsetty S, Tsoaras AM, Sha Y, Eissa NT, Hershey GK (2012) Functional variant in the autophagy-related 5 gene promoter is associated with childhood asthma. *PLoS One* 7:e333454, doi:10.1371/journal.pone.00333454
52. Poon AH, Chouiali F, Tse SM, Litonjua AA, Hussain SN, Baglolle CJ, Eidelman DH, Olivenstein R, Martin JG, Weiss ST, Hamid Q, Laprise C (2012) Genetic and histologic evidence for autophagy in asthma pathogenesis. *J Allergy Clin Immunol* 129:569–571, doi:10.1016/j.jaci.2011.09.035
53. Volkov A, Hagner S, Liser S, Alnahas S, Raifer H, Hellhund A, Garn H, Steinhoff U (2013)  $\beta$ 5i subunit deficiency of the immunoproteasome leads to reduced Th2 response in OVA induced acute asthma. *PLoS One* 8:e60565, doi:10.1371/journal.pone.0060565
54. Kenche H, Baty CJ, Vedagiri K, Shapiro SD, Blumental-Perry A (2013) Cigarette smoking affects oxidative protein folding in endoplasmic reticulum by modifying protein disulfide isomerase. *FASEB J* 27:965–977, doi:10.1096/fj.12-216234
55. Yao H, Rahman I (2011) Current concepts on oxidative/carbonyl stress, inflammation and epigenetics in pathogenesis of chronic obstructive pulmonary disease. *Toxicol Appl Pharmacol* 254:72–85, doi:10.1016/j.taap.2009.10.022
56. Ryter SW, Nakahira K, Haspel JA, Choi AMK (2012) Autophagy in pulmonary diseases. *Annu Rev Physiol* 74:377–401, doi:10.1146/annurev-physiol-020911-153348
57. Patel AS, Morse D, Choi AMK (2012) Regulation and functional significance of autophagy in respiratory cell biology and disease. *Am J Respir Cell Mol Biol*. doi:10.1165/rcmb.2012-0282TR
58. Dromparis P, Michelakis ED (2013) Mitochondria in vascular health and disease. *Annu Rev Physiol* 75:95–126, doi:10.1146/annurev-physiol-030212-183804
59. Patel AS, Lin L, Geyer A, Haspel JA, An CH, Cao J, Rosas IO, Morse D (2012) Autophagy in idiopathic pulmonary fibrosis. *PLoS One* 7:e41394, doi:10.1371/journal.pone.0041394
60. Baker TA, Bach HH 4th, Gamelli RL, Love RB, Majetschak M (2014) Proteasomes in lungs from organ donors and patients with end-stage pulmonary diseases. *Physiol. Res. Acad. Sci. Bohemoslov* 63(3):311–319
61. Malhotra D, Thimmulappa R, Navas-Acien A, Sandford A, Elliott M, Singh A, Chen L, Zhuang X, Hogg J, Pare P, Tudor RM, Biswal S (2008) Decline in NRF2-regulated antioxidants in chronic obstructive pulmonary disease lungs due to loss of its positive regulator, DJ-1. *Am J Respir Crit Care Med* 178:592–604, doi:10.1164/rccm.200803-3800C
62. Wei J, Rahman S, Ayaub EA, Dickhout JG, Ask K (2013) Protein misfolding and endoplasmic reticulum stress in chronic lung disease. *Chest J* 143:1098, doi:10.1378/chest.12-2133
63. Yeager ME, Reddy MB, Nguyen CM, Colvin KL, Ivy DD, Stenmark KR (2012) Activation of the unfolded protein response is associated with pulmonary hypertension. *Pulm Circ* 2:229–240
64. van Rijt SH, Keller IE, John G, Kohse K, Yildirim A7, Eickelberg O, Meiners S (2012) Acute cigarette smoke exposure impairs proteasome function in the lung. *Am J Physiol Lung Cell Mol Physiol* 303(9):L814–823
65. Somborac-Bacura A, van der Toorn M, Franciosi L, Slebos DJ, Zanic-Grubisic T, Bischoff R, van Oosterhout AJ (2012) Cigarette smoke induces endoplasmic reticulum stress response and proteasomal dysfunction in human alveolar epithelial cells. *Exp Physiol* 98:316–325, doi:10.1113/expphysiol.2012.067249
66. Lopez-Otin C, Blasco MA, Partridge L, Serrano M, Kroemer G (2013) The hallmarks of aging. *Cell* 153:1194–1217, doi:10.1016/j.cell.2013.05.039
67. Brown MK, Naidoo N (2012) The endoplasmic reticulum stress response in aging and age-related diseases. *Front Physiol* 3:263, doi:10.3389/fphys.2012.00263
68. Rojas M, Meiners S, Le Saux CJ (2014) Molecular Aspects of Aging: Understanding Lung Aging. John Wiley & Sons, Hoboken, New Jersey
69. Torres-Gonzalez E, Bueno M, Tanaka A, Krug LT, Cheng DS, Polosukhin WV, Sorescu D, Lawson WE, Blackwell TS, Rojas M, Mora AL (2012) Role of endoplasmic reticulum stress in age-related susceptibility to lung fibrosis. *Am J Respir Cell Mol Biol* 46:748–756, doi:10.1165/rcmb.2011-0224OC
70. Martinez J, Verbist K, Wang R, Green DR (2013) The relationship between metabolism and the autophagy machinery during the innate immune response. *Cell Metab* 17:895–900, doi:10.1016/j.cmet.2013.05.012
71. Osorio F, Lambrecht B, Janssens S (2013) The UPR and lung disease. *Semin Immunopathol* 35:293–306, doi:10.1007/s00281-013-0368-6

doi:10.1186/s40348-014-0011-1

**Cite this article as:** Meiners and Ballweg: Proteostasis in pediatric pulmonary pathology. *Molecular and Cellular Pediatrics* 2014 1:11.